

Pesticide-Induced Oxidative Stress and Neurodegeneration

Ivan Mutebi

Department of Pharmacognosy Kampala International University Uganda
Email: ivan.mutebi@studwc.kiu.ac.ug

ABSTRACT

Pesticides are widely used in agriculture and pest control, yet mounting evidence links chronic exposure to adverse effects on human health, particularly the nervous system. One key mechanism through which pesticides exert neurotoxicity is oxidative stress—an imbalance between pro-oxidant and antioxidant systems that results in cellular damage. Pesticide-induced oxidative stress triggers lipid peroxidation, DNA damage, mitochondrial dysfunction, and neuroinflammation, processes implicated in the pathogenesis of neurodegenerative diseases such as Parkinson's disease, Alzheimer's disease, amyotrophic lateral sclerosis, and Huntington's disease. This review synthesizes current understanding of how pesticides cause oxidative damage in neural tissues, examines cellular mechanisms involved, and explores epidemiological and experimental evidence linking pesticide exposure to neurodegeneration. We further evaluate protective strategies, the role of genetic susceptibility, and future research directions to mitigate pesticide-related neurotoxicity.

Keywords: Oxidative stress, Neurodegeneration, Pesticides, Mitochondrial dysfunction, Neuroinflammation

INTRODUCTION

Pesticides encompass a wide range of chemical compounds designed to control pests in agriculture and households, including insecticides, herbicides, fungicides, and rodenticides[1]. While beneficial in increasing crop yields, preserving stored food, and controlling vectors of infectious diseases, many pesticides are biologically active compounds that may adversely affect non-target organisms[2]. Their widespread and sometimes indiscriminate use has resulted in environmental persistence, bioaccumulation, and chronic low-level human exposure through contaminated food, water, soil, and air. Occupational exposure among agricultural workers further increases the risk of toxic effects. Over the past several decades, epidemiological and experimental studies have increasingly linked long-term pesticide exposure with neurological impairments and elevated risk of neurodegenerative disorders[3]. These associations are particularly concerning given the progressive and often irreversible nature of neurodegeneration. A central mechanism underlying pesticide-induced neurotoxicity is oxidative stress, a pathological condition characterized by excessive production of reactive oxygen species (ROS) and reactive nitrogen species (RNS) relative to endogenous antioxidant defenses[4]. When the balance between oxidants and antioxidants is disrupted, oxidative damage accumulates in cellular macromolecules, impairing neuronal structure and function. The brain is especially vulnerable to oxidative stress for several reasons. It consumes a disproportionately high amount of oxygen relative to its size, contains abundant polyunsaturated fatty acids that are highly susceptible to lipid peroxidation, and possesses relatively limited antioxidant reserves compared to other organs. In addition, neurons are post-mitotic cells with limited regenerative capacity, making oxidative injury particularly consequential. This review examines how pesticides promote oxidative stress in neural tissue, highlights key chemical classes associated with neurotoxicity, and outlines the cellular mechanisms that contribute to neuronal vulnerability and degeneration.

2. Pesticides Commonly Linked to Neurotoxicity

Pesticides represent diverse chemical classes with distinct molecular targets, yet many converge on common oxidative and neurotoxic pathways[5]. Among those most frequently implicated in neurodegenerative outcomes are organophosphates, organochlorines, pyrethroids, and certain herbicides such as paraquat and rotenone. Organophosphates, including chlorpyrifos and malathion, exert toxicity primarily through inhibition of acetylcholinesterase, leading to excessive accumulation of acetylcholine at synapses[6]. Beyond acute cholinergic

toxicity, chronic low-dose exposure has been shown to induce oxidative stress, disrupt mitochondrial integrity, and impair neurodevelopmental processes. Organochlorines such as DDT and lindane are highly lipophilic and resistant to degradation, enabling them to accumulate in adipose tissue and neural membranes[7]. Their persistence in the environment and bioaccumulative nature increase the likelihood of long-term exposure. These compounds interfere with ion channels and mitochondrial function, promoting ROS generation and membrane destabilization[8]. Pyrethroids, including permethrin, act on voltage-gated sodium channels to prolong neuronal excitation. Although generally considered less persistent than organochlorines, accumulating evidence indicates that pyrethroids can enhance oxidative stress indirectly through increased neuronal activity and secondary mitochondrial strain[9]. Herbicides such as paraquat and rotenone have drawn particular attention due to their strong experimental links to Parkinsonian pathology. Rotenone directly inhibits mitochondrial complex I of the electron transport chain, while paraquat undergoes redox cycling to produce superoxide radicals. Both mechanisms result in substantial ROS production and selective dopaminergic neuronal injury. Collectively, these pesticides increase oxidative burden either by stimulating ROS generation, impairing mitochondrial respiration, or weakening endogenous antioxidant defenses, thereby precipitating oxidative damage in neural tissues.

3. Oxidative Stress: Mechanisms and Neural Vulnerability

3.1 Reactive Oxygen and Nitrogen Species

Reactive oxygen species include free radicals such as superoxide and hydroxyl radicals, as well as non-radical oxidants like hydrogen peroxide[10]. Reactive nitrogen species, including nitric oxide and peroxynitrite, further contribute to oxidative and nitrosative stress. Under physiological conditions, ROS and RNS are generated as byproducts of mitochondrial respiration and participate in intracellular signaling[11]. However, excessive accumulation leads to oxidation of lipids, proteins, and nucleic acids, altering their structure and function.

3.2 Antioxidant Defenses in the Brain

Neural cells rely on enzymatic antioxidants such as superoxide dismutase, catalase, and glutathione peroxidase to neutralize ROS[12]. Non-enzymatic antioxidants, including glutathione and vitamins C and E, provide additional protection. Despite these systems, the brain's antioxidant capacity is comparatively modest, rendering it susceptible to oxidative insults, particularly during chronic toxicant exposure.

3.3 Mitochondria as Targets

Mitochondria are both a primary source and a major target of ROS. Pesticides such as rotenone disrupt the electron transport chain, causing electron leakage and enhanced ROS production[13]. Damaged mitochondria generate less ATP, impair calcium homeostasis, and activate apoptotic signaling pathways. In neurons, which depend heavily on aerobic metabolism, mitochondrial dysfunction can rapidly compromise synaptic transmission and cell survival, amplifying the impact of oxidative stress[14].

4. Pathways of Pesticide-Induced Oxidative Stress

Pesticides induce oxidative stress through interconnected molecular and cellular mechanisms that ultimately compromise neuronal survival[15]. These processes are not isolated events but part of a complex cascade involving membrane damage, genomic instability, protein dysfunction, mitochondrial impairment, and chronic inflammation[16]. The cumulative impact of these alterations contributes significantly to progressive neurodegeneration.

4.1 Lipid Peroxidation

Reactive oxygen species readily attack polyunsaturated fatty acids within neuronal plasma membranes and intracellular organelle membranes, initiating lipid peroxidation[17]. This chain reaction generates highly reactive aldehydes such as malondialdehyde and 4-hydroxynonenal, which further propagate oxidative injury by forming adducts with proteins and DNA. Because neuronal membranes are rich in polyunsaturated lipids, particularly in synaptic terminals and myelin sheaths, they are especially susceptible to peroxidative damage[18]. Lipid peroxidation disrupts membrane fluidity and permeability, altering ion gradients essential for action potential generation and neurotransmitter release. Damage to synaptic membranes impairs vesicle fusion and receptor function, leading to defective synaptic transmission. In addition, oxidation of mitochondrial membranes compromises electron transport chain integrity, increasing electron leakage and further ROS production[19]. Thus, lipid peroxidation both results from and amplifies oxidative stress, creating a self-perpetuating cycle of neuronal injury.

4.2 DNA and Protein Damage

Oxidative stress also targets nuclear and mitochondrial DNA. ROS can induce base modifications, single- and double-strand breaks, and cross-linking[20]. One commonly observed lesion is 8-hydroxy-2'-deoxyguanosine, a marker of oxidative DNA damage. If not efficiently repaired, such lesions may lead to mutations, impaired transcription, and defective mitochondrial protein synthesis. Because neurons are long-lived cells with limited regenerative capacity, accumulated DNA damage can progressively impair cellular function[21]. Proteins are similarly vulnerable to oxidative modification. Oxidation of amino acid side chains, formation of carbonyl groups, and disruption of disulfide bonds can alter protein conformation and enzymatic activity. Key neuronal proteins,

including cytoskeletal components and metabolic enzymes, may lose structural integrity or become prone to aggregation[22]. Impairment of proteasomal and autophagic degradation pathways further exacerbates accumulation of damaged proteins. Over time, these molecular alterations compromise synaptic maintenance, axonal transport, and neuronal survival.

4.3 Neuroinflammation

Oxidative stress activates resident immune cells of the central nervous system, particularly microglia and astrocytes[23]. Upon activation, microglia release pro-inflammatory cytokines such as tumor necrosis factor-alpha and interleukin-1 beta, as well as additional reactive oxygen and nitrogen species. Astrocytes may also contribute by producing inflammatory mediators and altering glutamate homeostasis[24]. While acute neuroinflammation can serve protective functions, chronic activation becomes detrimental. Sustained inflammatory signaling enhances oxidative stress, disrupts the blood-brain barrier, and promotes excitotoxicity through excessive glutamate release[25]. This persistent inflammatory environment amplifies neuronal injury and establishes a vicious cycle in which oxidative stress and inflammation mutually reinforce each other. Such chronic neuroinflammation is increasingly recognized as a key contributor to progressive neurodegenerative processes associated with pesticide exposure.

5. Pesticide Exposure and Neurodegenerative Diseases

Growing epidemiological and experimental evidence links pesticide-induced oxidative stress with major neurodegenerative disorders[26-30]. Although causality is complex and multifactorial, oxidative mechanisms provide a biologically plausible bridge between environmental exposure and neuronal degeneration.

5.1 Parkinson's Disease (PD)

Parkinson's disease is characterized by progressive degeneration of dopaminergic neurons in the substantia nigra pars compacta and accumulation of misfolded alpha-synuclein aggregates[27,31-34]. Dopaminergic neurons are particularly susceptible to oxidative stress due to dopamine metabolism, which itself generates reactive intermediates[28,35-38]. Epidemiological studies have consistently reported increased Parkinson's disease risk among individuals exposed to pesticides such as paraquat and rotenone. Rotenone inhibits mitochondrial complex I, impairing ATP production and promoting excessive ROS generation. Experimental models using rotenone replicate key features of Parkinsonian pathology, including dopaminergic neuron loss and motor deficits[29,39-42]. Paraquat, through redox cycling, produces superoxide radicals that selectively damage dopaminergic neurons. Oxidative stress also promotes alpha-synuclein misfolding and aggregation, further linking pesticide exposure to Parkinson's disease pathogenesis.

5.2 Alzheimer's Disease (AD)

Alzheimer's disease is marked by extracellular amyloid-beta plaques, intracellular neurofibrillary tangles composed of hyperphosphorylated tau, and widespread synaptic loss[30,43-46]. Oxidative stress is an early and prominent feature of Alzheimer's pathology. Chronic exposure to certain pesticides has been associated with cognitive decline and increased Alzheimer's disease risk in some population studies[31,47-50]. Organophosphates such as chlorpyrifos can enhance oxidative stress in cortical and hippocampal neurons, regions critical for memory and learning. Oxidative damage may accelerate amyloid-beta aggregation and tau phosphorylation, while mitochondrial dysfunction impairs neuronal energy metabolism[32,50-51]. These mechanisms suggest that pesticide-induced oxidative stress may contribute to both the initiation and progression of Alzheimer's disease.

5.3 Amyotrophic Lateral Sclerosis (ALS)

Amyotrophic lateral sclerosis involves degeneration of upper and lower motor neurons, leading to progressive muscle weakness and paralysis[33]. Oxidative stress and excitotoxicity are central features of ALS pathophysiology. Some occupational studies indicate elevated ALS incidence among agricultural workers exposed to pesticides, although results vary across populations. Pesticide-induced oxidative damage may exacerbate glutamate-mediated excitotoxicity and impair mitochondrial function in motor neurons[34]. Given that mutations in antioxidant-related genes are implicated in familial ALS, environmental oxidative stressors such as pesticides may act synergistically with genetic predispositions to increase disease risk.

5.4 Huntington's Disease (HD)

Huntington's disease is a hereditary neurodegenerative disorder caused by expanded CAG repeats in the huntingtin gene[35]. Although primarily genetic in origin, oxidative stress plays a significant role in neuronal dysfunction and striatal degeneration associated with the disease. Environmental toxicants, including pesticides, may intensify oxidative burden and mitochondrial impairment in vulnerable neurons[36]. Exposure to pro-oxidant pesticides could potentially accelerate symptom onset or worsen disease severity by compounding existing mitochondrial deficits and promoting protein aggregation[37]. While further research is needed, oxidative stress provides a plausible mechanistic link between environmental exposures and modification of Huntington's disease progression.

CONCLUSION

Pesticides contribute to oxidative stress in the nervous system through multiple mechanisms, including ROS overproduction, mitochondrial dysfunction, lipid peroxidation, and neuroinflammation. Evidence from cellular models, animal studies, and human research supports links between pesticide exposure and neurodegenerative diseases. While reducing exposure remains primary prevention, ongoing research into antioxidants, genetic susceptibility, and safer alternatives is vital. Protecting neurological health demands coordinated effort from scientists, clinicians, policymakers, and communities.

REFERENCES

1. Felicia N Ekeh, Gregory E Odo, Ekechukwu Nkiru, EJ Agwu, Clara Ikegbunam, S Haruna Ada (2015). Effects of biopesticides on developmental stages and longevity of *Callosobruchus maculatus* in some leguminous grains. *Journal of Parasitology and Vector Biology*, 7, (1), 9-21.
2. Tudi M, Daniel Ruan H, Wang L, Lyu J, Sadler R, Connell D, Chu C, Phung DT. Agriculture Development, Pesticide Application and Its Impact on the Environment. *International Journal of Environmental Research and Public Health*. 2021; 18(3):1112. <https://doi.org/10.3390/ijerph18031112>
3. Egba, Simeon Ikechukwu; Okonkwo, Chibuzor Onyinye; Kalu, Uguru Grace Paul Promise, Chibuike, Sampson Nzubechi L, Egbachukwu Blessing Ifeoma, Alum Esther Ugo and Ugwu Okechukwu Paul-Chima (2024). Ameliorative potentials of methanol leaf extract of *Anthocleista vogelli* on mercury chloride induced neurotoxicity. *INOSR Experimental Sciences* 13(2):15-22. <https://doi.org/10.59298/INOSRES/2024/1321522.000>
4. Egba, S Ikechukwu, Okonkwo C Onyinye, Ogbodo J Onyebuchi and Ezech V Nzubechukwu Neuroprotective Potentials of *Alstonia boonei* extracts on Biochemical Markers of Brain Integrity in Experimental Rats, *Trop J Nat Prod Res*, 2021; 5(6): 1106-1109. doi.org/10.26538/tjnpr/v5i6.21
5. Muñoz-Bautista JM, Bernal-Mercado AT, Martínez-Cruz O, Burgos-Hernández A, López-Zavala AA, Ruiz-Cruz S, Ornelas-Paz JdJ, Borboa-Flores J, Ramos-Enríquez JR, Del-Toro-Sánchez CL. Environmental and Health Impacts of Pesticides and Nanotechnology as an Alternative in Agriculture. *Agronomy*. 2025; 15(8):1878. <https://doi.org/10.3390/agronomy15081878>
6. Egba, S.I., Ademola C F and Omoruyi L E (2021) Bucholzia coriacea seed extract attenuates mercury induced cerebral and cerebellar oxidative neurotoxicity via NO signalling and suppression of oxidative stress, adenosine deaminase and acetylcholinesterase activities in rats. *Avicenna J Phytomed*. 2022 Jan-Feb;12(1):42-53. doi: 10.22038/AJP.2021.18262. PMID: 35145894; PMCID: PMC8801217.
7. Petrovici A, Savuța G, Lucini C, Robea M-A, Solcan C. Combined Neurotoxic Effects of Commercial Formulations of Pyrethroid (Deltamethrin) and Neonicotinoid (Imidacloprid) Pesticides on Adult Zebrafish (*Danio rerio*): Behavioral, Molecular, and Histopathological Analysis. *Life*. 2025; 15(4):538. <https://doi.org/10.3390/life15040538>
8. Naima J, Ohta Y. Potassium Ions Decrease Mitochondrial Matrix pH: Implications for ATP Production and Reactive Oxygen Species Generation. *Int J Mol Sci*. 2024 Jan 19;25(2):1233. doi: 10.3390/ijms25021233. PMID: 38279231; PMCID: PMC10815940.
9. Zorov DB, Juhaszova M, Sollott SJ. Mitochondrial reactive oxygen species (ROS) and ROS-induced ROS release. *Physiol Rev*. 2014 Jul;94(3):909-50. doi: 10.1152/physrev.00026.2013. PMID: 24987008; PMCID: PMC4101632.
10. Uhwo E N, Egba S I, Nwuke P C, Obike C A and Kelechi G K. Antioxidative properties of *Adansonia digitata* L. (baobab) leaf extract exert protective effect on doxorubicin induced cardiac toxicity in Wistar rats. *Clinical Nutrition Open Science* 2022; 45:3-16
11. Ugwu, CE., Sure, SM., Dike, CC., Okpoga, NA and Egba, SI. Phytochemical and *in vitro* antioxidant activities of methanol leave extract of *Alternanthera basiliana*. *Journal of Pharmacy Research*, 2018; 12(6): 835-839
12. Alum, E.U., Uti, D.E. & Offor, C.E. Redox Signaling Disruption and Antioxidants in Toxicology: From Precision Therapy to Potential Hazards. *Cell Biochem Biophys* (2025). <https://doi.org/10.1007/s12013-025-01846-8>
13. Giorgi C, Marchi S, Simoes ICM, Ren Z, Morciano G, Perrone M, Patalas-Krawczyk P, Borchard S, Jędrak P, Pierzynowska K, Szymański J, Wang DQ, Portincasa P, Węgrzyn G, Zischka H, Dobrzym P, Bonora M, Duszynski J, Rimessi A, Karkucinska-Wieckowska A, Dobrzym A, Szabadkai G, Zavan B, Oliveira PJ, Sardao VA, Pinton P, Wieckowski MR. Mitochondria and Reactive Oxygen Species in Aging and Age-Related Diseases. *Int Rev Cell Mol Biol*. 2018;340:209-344. doi: 10.1016/bs.ircmb.2018.05.006. Epub 2018 Jun 22. PMID: 30072092; PMCID: PMC8127332.
14. Ogwoni, H. A., Aja, P. M., Eze, E. D., Agu, P. C., Moyosore, A. A., Ale, B. A., ... & Awuchi, C. G. (2024). *Cucumeropsis mannii* seed oil (CMSO) restores testicular mitochondrial dysfunctions by modulating the

- activities of dysregulated testicular mitochondrial enzymes in male albino rats exposed to bisphenol A. *Food Science & Nutrition*, 12(10), 7854. doi: 10.1002/fsn3.4379. PMID: 39479659; PMCID: PMC11521674.
15. Sule RO, Condon L, Gomes AV. A Common Feature of Pesticides: Oxidative Stress-The Role of Oxidative Stress in Pesticide-Induced Toxicity. *Oxid Med Cell Longev*. 2022 Jan 19;2022:5563759. doi: 10.1155/2022/5563759. PMID: 35096268; PMCID: PMC8791758.
 16. Ugwu, O.P., Okon, M.B., Alum, E.U., Ugwu, C.N., Anyanwu, E.G., Mariam, B., et al. (2025). Unveiling the therapeutic potential of the gut microbiota-brain axis: Novel insights and clinical applications in neurological disorders. *Medicine (Baltimore)*. 2025 Jul 25;104(30):e43542. doi: 10.1097/MD.00000000000043542. PMID: 40725913; PMCID: PMC12303509.
 17. Su LJ, Zhang JH, Gomez H, Murugan R, Hong X, Xu D, Jiang F, Peng ZY. Reactive Oxygen Species-Induced Lipid Peroxidation in Apoptosis, Autophagy, and Ferroptosis. *Oxid Med Cell Longev*. 2019 Oct 13;2019:5080843. doi: 10.1155/2019/5080843. PMID: 31737171; PMCID: PMC6815535.
 18. Ayala A, Muñoz MF, Argüelles S. Lipid peroxidation: production, metabolism, and signaling mechanisms of malondialdehyde and 4-hydroxy-2-nonenal. *Oxid Med Cell Longev*. 2014;2014:360438. doi: 10.1155/2014/360438. Epub 2014 May 8. PMID: 24999379; PMCID: PMC4066722.
 19. Juan CA, Pérez de la Lastra JM, Plou FJ, Pérez-Lebeña E. The Chemistry of Reactive Oxygen Species (ROS) Revisited: Outlining Their Role in Biological Macromolecules (DNA, Lipids and Proteins) and Induced Pathologies. *International Journal of Molecular Sciences*. 2021; 22(9):4642. <https://doi.org/10.3390/ijms22094642>
 20. Han Y, Chen JZ. Oxidative stress induces mitochondrial DNA damage and cytotoxicity through independent mechanisms in human cancer cells. *Biomed Res Int*. 2013;2013:825065. doi: 10.1155/2013/825065. Epub 2012 Dec 26. PMID: 23509785; PMCID: PMC3591153.
 21. Tripathi D, Oldenburg DJ, Bendich AJ. Oxidative and Glycation Damage to Mitochondrial DNA and Plastid DNA during Plant Development. *Antioxidants*. 2023; 12(4):891. <https://doi.org/10.3390/antiox12040891>
 22. Pizzino G, Irrera N, Cucinotta M, Pallio G, Mannino F, Arcoraci V, Squadrito F, Altavilla D, Bitto A. Oxidative Stress: Harms and Benefits for Human Health. *Oxid Med Cell Longev*. 2017;2017:8416763. doi: 10.1155/2017/8416763. Epub 2017 Jul 27. PMID: 28819546; PMCID: PMC5551541.
 23. Chen Y, Qin C, Huang J, Tang X, Liu C, Huang K, Xu J, Guo G, Tong A, Zhou L. The role of astrocytes in oxidative stress of central nervous system: A mixed blessing. *Cell Prolif*. 2020 Mar;53(3):e12781. doi: 10.1111/cpr.12781. Epub 2020 Feb 8. PMID: 32035016; PMCID: PMC7106951.
 24. Grasso M, Mascali C, L'Episcopo F. Role of Reactive Astrocytes and Microglia: Wnt/ β -Catenin Signaling in Neuroprotection and Repair in Parkinson's Disease. *International Journal of Molecular Sciences*. 2025; 26(24):11880. <https://doi.org/10.3390/ijms262411880>
 25. de Araújo AB, S Azul FVC, Carneiro YC, de Sousa CNS, de Vasconcelos SMM, Rios FJ, Leal LKAM. Neuroinflammation and Oxidative Stress in Parkinson's Disease, Alzheimer's Disease, and COVID-19: Microglia-Neutrophil Interaction. *ACS Omega*. 2026 Jan 26;11(5):6922-6938. doi: 10.1021/acsomega.5c10397. PMID: 41696228; PMCID: PMC12902863.
 26. Rodríguez A, Castrejón-Godínez ML, Monterrosas-Brisson N. Pesticides: Environmental Stressors Implicated in the Development of Central Nervous System Disorders and Neurodegeneration. *Stresses*. 2025; 5(2):31. <https://doi.org/10.3390/stresses5020031>
 27. Srinivasan E, Chandrasekhar G, Chandrasekar P, Anbarasu K, Vickram AS, Karunakaran R, Rajasekaran R, Srikumar PS. Alpha-Synuclein Aggregation in Parkinson's Disease. *Front Med (Lausanne)*. 2021 Oct 18;8:736978. doi: 10.3389/fmed.2021.736978. PMID: 34733860; PMCID: PMC8558257.
 28. Pitton Rissardo J, McGarry A, Shi Y, Fornari Caprara AL, Kannarkat GT. Alpha-Synuclein Neurobiology in Parkinson's Disease: A Comprehensive Review of Its Role, Mechanisms, and Therapeutic Perspectives. *Brain Sciences*. 2025; 15(12):1260. <https://doi.org/10.3390/brainsci15121260>
 29. Kouli A, Torsney KM, Kuan WL. Parkinson's Disease: Etiology, Neuropathology, and Pathogenesis. In: Stoker TB, Greenland JC, editors. *Parkinson's Disease: Pathogenesis and Clinical Aspects* [Internet]. Brisbane (AU): Codon Publications; 2018 Dec 21. Chapter 1. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK536722/> doi: 10.15586/codonpublications.parkinsonsdisease.2018.ch1
 30. Tenchov R, Sasso JM, Zhou QA. Alzheimer's Disease: Exploring the Landscape of Cognitive Decline. *ACS Chem Neurosci*. 2024 Nov 6;15(21):3800-3827. doi: 10.1021/acscchemneuro.4c00339. Epub 2024 Oct 11. PMID: 39392435; PMCID: PMC11587518.
 31. Uti DE, Egba SI, Ugwu OP-C, Aja PM. The Role of Phytochemicals in Age-Related Cognitive Decline: A Natural Solution for Brain Health. *Natural Product Communications*. 2025;20(6).

- doi:10.1177/1934578X251350761
32. Zhang H, Wei W, Zhao M, Ma L, Jiang X, Pei H, Cao Y, Li H. Interaction between A β and Tau in the Pathogenesis of Alzheimer's Disease. *Int J Biol Sci.* 2021 May 27;17(9):2181-2192. doi: 10.7150/ijbs.57078. PMID: 34239348; PMCID: PMC8241728.
 33. Brotman RG, Moreno-Escobar MC, Joseph J, et al. Amyotrophic Lateral Sclerosis. [Updated 2024 Feb 12]. In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishing; 2025 Jan-. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK556151/>
 34. Ajitkumar A, Lui F, De Jesus O. Huntington Disease. [Updated 2025 Apr 6]. In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishing; 2025 Jan-. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK559166/>
 35. Ross CA, Tabrizi SJ. Huntington's disease: from molecular pathogenesis to clinical treatment. *Lancet Neurol.* 2011 Jan;10(1):83-98. doi: 10.1016/S1474-4422(10)70245-3. PMID: 21163446.
 36. Tong H, Yang T, Xu S, Li X, Liu L, Zhou G, Yang S, Yin S, Li X-J, Li S. Huntington's Disease: Complex Pathogenesis and Therapeutic Strategies. *International Journal of Molecular Sciences.* 2024; 25(7):3845. <https://doi.org/10.3390/ijms25073845>
 37. Ugwu CN, Ugwu OPC, Alum EU, Eze VHU, Basajja M, Ugwu JN, et al. Medical preparedness for bioterrorism and chemical warfare: a public health integration review. *Medicine.* 2025;104(18):e42289.
 38. Ugwu CN, Ugwu OPC, Alum EU, Eze VHU, Basajja M, Ugwu JN, et al. Sustainable development goals (SDGs) and resilient healthcare systems: addressing medicine and public health challenges in conflict zones. *Medicine.* 2025;104(7):e41535.
 39. Ugwu OPC, Alum EU, Ugwu JN, Eze VHU, Ugwu CN, Ogenyi FC, et al. Harnessing technology for infectious disease response in conflict zones: challenges, innovations, and policy implications. *Medicine.* 2024;103(28):e38834.
 40. Nyamboga TO, Ugwu OPC, Ugwu CN, Ogenyi FC, Alum EU, Ugwu JN. Optimizing emergency response systems in urban health crises: a project management approach to public health preparedness and response. *Medicine.* 2025;104(3):e41279.
 41. Alum EU, Obeagu EI, Ugwu OPC. Enhancing quality water, good sanitation, and proper hygiene is the panacea to diarrhea control and the attainment of some related sustainable development goals: a review. *Medicine.* 2024;103(38):e39578.
 42. Alum EU, Obeagu EI, Ugwu OPC. Curbing diarrhea in children below five years old: the sub-Saharan African standpoint. *J New Med Innov Res.* 2024;5(1).
 43. Ezenwaji CO, Alum EU, Ugwu OPC. The role of digital health in pandemic preparedness and response: securing global health? *Glob Health Action.* 2024;17(1):2419694.
 44. Edyedu I, Ugwu OPC, Ugwu CN, Alum EU, Aja PM. The role of pharmacological interventions in managing urological complications during pregnancy and childbirth: a review. *Medicine.* 2025;104(7):e41381.
 45. Anyanwu GE, Ejemot-Nwadiaro RI, Ojiakor OV, Makena W, Ugwu OPC, Alum EU, et al. Teaching through chaos: resilience and survival of female educators in refugee and conflict-affected settings. *F1000Res.* 2026;15:41.
 46. Alum EU, Ugwu OPC, Obeagu EI, Okon MB. Religious leaders as advocates for promoting exclusive breastfeeding in East Africa. *Int J Innov Appl Res.* 2023;11(12):10-5.
 47. Alum EU, Obeagu EI, Ugwu OPC, Ugwu CN, Uti DE, Samson AO, et al. Nutritional requirements during pregnancy: a comprehensive overview. *Int J Innov Appl Res.* 2023;11(12):26-34.
 48. Obeagu EI, Obeagu GU, Igwe MC, Alum EU, Ugwu OPC. Neutrophil-derived inflammation and pregnancy outcomes. *Newport Int J Sci Exp Sci.* 2023;4(2).
 49. Alum EU, Obeagu EI, Ugwu OPC, Uti DE, Alum BN, Ugwu CN. Mental health interventions for pregnant and postpartum women: efficacy and accessibility. *Elite J Nurs Health Sci.* 2024;2(6):43-9.
 50. Ugwu OPC, Ugwu CN, Ogenyi FC. Innovations in renewable energy for health applications. *Eurasian Exp J Med Med Sci.* 2025;6(1):75-80.
 51. Ogutu V, Jacob EE, Vahwere BM, Oviosun A, Anyanwu CN, Makena W, et al. Global prevalence, patterns, and predictors of relapse following Ponseti treatment for idiopathic clubfoot: a systematic review and meta-analysis. *BMC Musculoskelet Disord.* 2026.

CITE AS: Ivan Mutebi. (2026). Pesticide-Induced Oxidative Stress and Neurodegeneration. IDOSR JOURNAL OF SCIENCE AND TECHNOLOGY 12(2):15-21, 2026.
<https://doi.org/10.59298/IDOSR/JST/26/122.1521>